

Connecting via Winsock to STN

Welcome to STN International! Enter x:x

LOGINID: ssspta1653rbm

PASSWORD:

TERMINAL (ENTER 1, 2, 3, OR ?):2

Enter NEWS followed by the item number or name to see news on that specific topic.


```
2 FILE SCISEARCH
1 FILE TOXCENTER
203 FILE USPATFULL
22 FILE USPAT2
1 FILE WATER
72 FILES SEARCHED...
```

```
18 FILES HAVE ONE OR MORE ANSWERS, 75 FILES SEARCHED IN STNINDEX
```

```
L1 QUE SEQUENCING AND DIAZOMETHANE
```

```
=> d rank
F1      203  USPATFULL
F2      22   USPAT2
F3      7    CAPLUS
F4      4    MEDLINE
F5      2    BIOSIS
F6      2    IFIPAT
F7      2    SCISEARCH
F8      1    AGRICOLA
F9      1    ANABSTR
F10     1    BIOTECHNO
F11     1    CABA
F12     1    CEN
F13     1    DISSABS
F14     1    EMBASE
F15     1    PASCAL
F16     1    PROMT
F17     1    TOXCENTER
F18     1    WATER
```

```
=> file caplus medline biosis scisearch
COST IN U.S. DOLLARS          SINCE FILE        TOTAL
                                ENTRY           SESSION
FULL ESTIMATED COST          2.36            5.52
```

```
FILE 'CAPLUS' ENTERED AT 14:49:00 ON 09 MAR 2005 .
USE IS SUBJECT TO THE TERMS OF YOUR STN CUSTOMER AGREEMENT.
PLEASE SEE "HELP USAGETERMS" FOR DETAILS.
COPYRIGHT (C) 2005 AMERICAN CHEMICAL SOCIETY (ACS)
```

```
FILE 'MEDLINE' ENTERED AT 14:49:00 ON 09 MAR 2005
```

```
FILE 'BIOSIS' ENTERED AT 14:49:00 ON 09 MAR 2005
Copyright (c) 2005 The Thomson Corporation
```

```
FILE 'SCISEARCH' ENTERED AT 14:49:00 ON 09 MAR 2005
Copyright (c) 2005 The Thomson Corporation
```

```
=> sequencing and diazomethane
L2      15 SEQUENCING AND DIAZOMETHANE
```

```
=> dup remove
ENTER L# LIST OR (END):12
PROCESSING COMPLETED FOR L2
L3      8 DUP REMOVE L2 (7 DUPLICATES REMOVED)
```

```
=> d ti 1-8
```

```
L3  ANSWER 1 OF 8 CAPLUS COPYRIGHT 2005 ACS on STN DUPLICATE 1
TI  Development of an analytical scheme for simazine and 2,4-D in soil and
    water runoff from ornamental plant nursery plots
```

L3 ANSWER 2 OF 8 CAPLUS COPYRIGHT 2005 ACS on STN
TI Purification and partial amino acid sequences of an esterase from tomato

L3 ANSWER 3 OF 8 CAPLUS COPYRIGHT 2005 ACS on STN DUPLICATE 2
TI Cathepsin B-like cysteine proteases confer intestinal cysteine protease activity in *Haemonchus contortus*

L3 ANSWER 4 OF 8 CAPLUS COPYRIGHT 2005 ACS on STN
TI Covalent modification of 2'-hydroxyl groups of RNA

L3 ANSWER 5 OF 8 CAPLUS COPYRIGHT 2005 ACS on STN DUPLICATE 3
TI Processing the procarboxypeptidase A and other zymogens in murine mast cells

L3 ANSWER 6 OF 8 MEDLINE on STN
TI The catecholamine binding site of the beta-adrenergic receptor is formed by juxtaposed membrane-spanning domains.

L3 ANSWER 7 OF 8 CAPLUS COPYRIGHT 2005 ACS on STN
TI Specific termination of RNA polymerase synthesis as a method of RNA and DNA sequencing

L3 ANSWER 8 OF 8 CAPLUS COPYRIGHT 2005 ACS on STN
TI Peptide sequencing by low-resolution mass spectrometry. I. Use of Acetylacetonyl derivatives to identify N-terminal residues

=> d ab bib 2, 8

L3 ANSWER 2 OF 8 CAPLUS COPYRIGHT 2005 ACS on STN
AB Screening of 18 suspension plant cell cultures of taxonomically distant species revealed that a Me jasmonate hydrolyzing enzyme activity (0.21-5.67 pkat/mg) occurs in all species so far analyzed. The Me jasmonate hydrolyzing esterase was purified from cell cultures of *Lycopersicon esculentum* using a five-step procedure including anion-exchange chromatog., gel-filtration and chromatog. on hydroxylapatite. The esterase was purified 767-fold to give an almost homogeneous protein in a yield of 2.2%. The native enzyme exhibited a Mr of 26 kDa (gel-filtration chromatog.), which was similar to the Mr determined by SDS-PAGE and MALDI-TOF anal. (Mr of 28547 kDa). Enzyme kinetics revealed a Km value of 15 μ M and a Vmax value of 7.97 nkat/mg, an pH optimum of 9.0 and a temperature optimum of 40 °C. The enzyme also efficiently hydrolyzed Me esters of abscisic acid, indole-3-acetic acid, and fatty acids. In contrast, Me esters of salicylic acid, benzoic acid and cinnamic acid were only poor substrates for the enzyme. N-Methylmaleimide, iodoacetamide, bestatin and pepstatin (inhibitors of thiol-, metal- and carboxyproteases, resp.) did not inactivate the enzyme while a serine protease inhibitor, phenylmethylsulfonyl fluoride, at a concentration of 5 mM led to irreversible and complete inhibition of enzyme activity. Proteolysis of the pure enzyme with endoproteinase LysC revealed three peptide fragments with 11-14 amino acids. N-Terminal sequencing yielded an addnl. peptide fragment with 10 amino acids. Sequence alignment of these fragments showed high homologies to certain plant esterases and hydroxynitrile lyases that belong to the α/β hydrolase fold protein superfamily.

AN 2002:382784 CAPLUS
DN 138:85391
TI Purification and partial amino acid sequences of an esterase from tomato
AU Stuhlfelder, Christiane; Lottspeich, Friedrich; Mueller, Martin J.
CS Julius-von-Sachs-Institute for Biosciences, Pharmaceutical Biology,
University of Wurzburg, Wurzburg, D-97082, Germany
SO Phytochemistry (2002), 60(3), 233-240

CODEN: PYTCAS; ISSN: 0031-9422
PB Elsevier Science Ltd.
DT Journal
LA English
RE.CNT 19 THERE ARE 19 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L3 ANSWER 8 OF 8 CAPLUS COPYRIGHT 2005 ACS on STN
AB A description was given of a low-resolution, mass-spectrometric method for the sequencing of acetylacetonyl (ACA) peptides (I) which gave reliable results with small (2-10 amino acid residues) I, regardless of the amino acids present, and depending on the identification, in the mass spectra visualized, of the N-terminal amino acid residue, "A," which has been found to represent the most prominent peak in the spectra of ACA I esters with N-terminal aliphatic or acidic amino acids. The prominence of these "A" ions in the system employed afforded an unambiguous starting point in the search for the sequence ions (B, C, D, and B1, C1, D1). Under the exptl. conditions employed, arginine I were converted to 8-N-(2-pyrimidinyl) ornithine I, and the ϵ -amino group of lysine was also derivatized; all other functional groups present in the protein amino acids remained intact and were left unprotected. In some cases, partial loss of the side chain was observed with N-terminal methionine, serine, threonine, aspartic acid, and glutamic acid. Some larger seryl- and threonyl-I tended to dehydrate at higher probe temps., making it difficult to recognize these residues in the N-terminal position. Histidyl-, tyrosyl-, phenylalanyl-, and tryptophyl-I showed some elimination of the side chain as ArCH2+ or ArCH2O, but these ions helped to confirm the presence of these residues. Lysyl- and arginyl-I yielded very characteristic [A-99] fragments, and showed only very small "A" fragments in the mass spectra, while cystine derivs. underwent SS bond rupture, accompanied by H+ transfer. Although I containing unmodified asparagine have been sequenced, diazomethane reportedly has to be used instead of alc. HCl for esterification, to prevent hydrolysis to the corresponding aspartyl-I. The ACA-I esters have a relatively high vapor pressure and yielded readily interpretable mass spectra from hepta- and octa-I containing only the neutral amino acids. The presence of basic, polyfunctional amino acids decreased the volatility and limited the sequence procedure to tetra- and penta-I. Expts. to increase the volatility of the ACA-I esters by permethylation reportedly invariably yielded a complex mixture of products, the potential difficulty being the enamino ketone function, which reacts with MeI to give O and C alkylation and a nonvolatile quaternary salt.

AN 1970:51653 CAPLUS
DN 72:51653
TI Peptide sequencing by low-resolution mass spectrometry. I. Use of Acetylacetonyl derivatives to identify N-terminal residues
AU Bacon, V.; Jellum, E.; Patton, W.; Pereira, W.; Halpern, B.
CS Med. Center, Stanford Univ., Stanford, CA, USA
SO Biochemical and Biophysical Research Communications (1969), 37(6), 878-82
CODEN: BBRCA9; ISSN: 0006-291X
DT Journal
LA English